

METHODS FOR THE ISOLATION OF DNA

- Salting out/salt precipitation method
- Phenol/chloroform extraction method
- Silica gel extraction method
- Proteinase K extraction method
- Anion exchange methos



The choice of method depends on many factors:

- Required quantity
- Purity of DNA
- Time
- Molecular weight of DNA

Equipment/reagent requirements

Red blood cell lysis buffer

White blood cell lysis buffer

RNAase

Protein precipitation solution

Isopropanol

DNA hydration buffer

A centrifuge machine

A vortex mixer

-80 degree Freezer

Incubator/Water-bath

A spectrophotometer capable of reading at 260 and 280 nm/Nanodrop



Choice of specimen

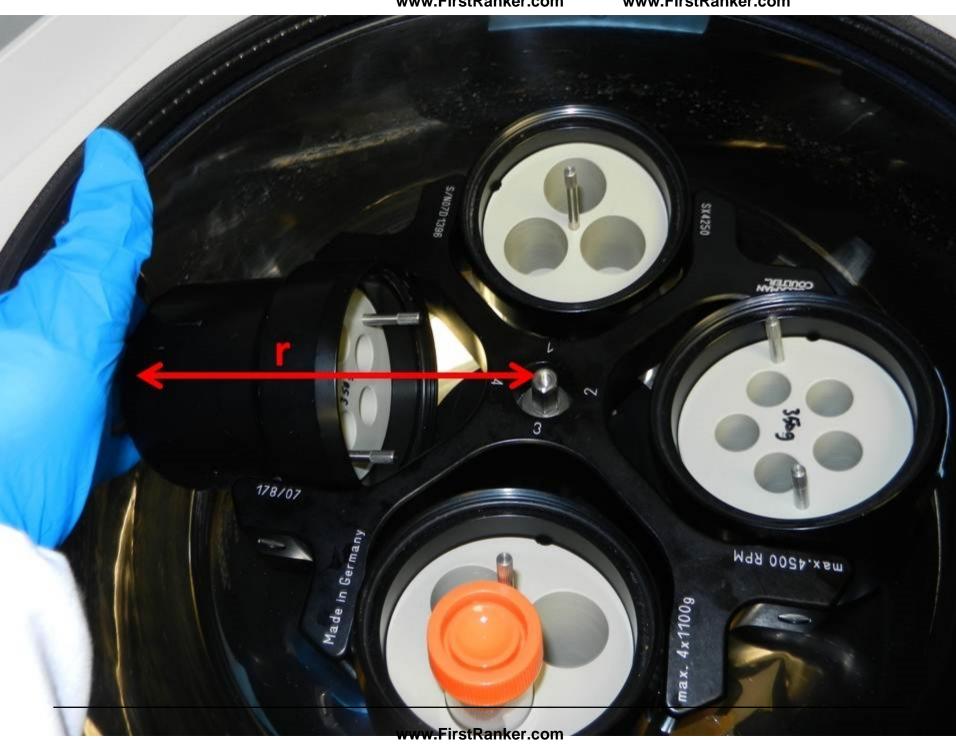
 DNA can be isolated from whole blood (EDTA tube) or a cell pellet following plasma separation from blood collected in EDTA tube

 Specimen can be stored at 4 degree Celsius for 48 hours prior to processing.

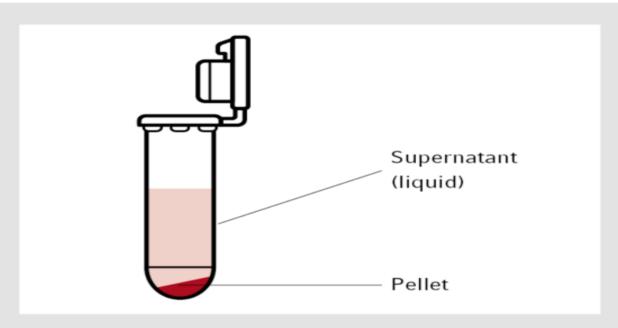


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Precipitate (pellet) stands for the concentrated particles in a tube after successful centrifugation.

Supernatant is the remaining solution above the pellet.



Procedure

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Cell lysis

- 1. Dispense 30ml of red blood cell lysis buffer (NH4CL, NaHCO3, EDTA) in to a 50 ml centrifuge tube containing 5-10 ml of whole blood. Incubate at RT for 5 min, inverting occasionally to mix.
- 2. Centrifuge the samples at 3,000g for to pellet the white blood cells. Pour the supernatant to waste.
- 3. Add 10ml of white cell lysis buffer (SDS, EDTA) to white blood cell pellet and vortex vigorously for 10 sec.
- 4. RNAase is added to remove RNA from the preparation .Incubate at 37 degree Celsius for 15 minutes.



Protein Precipitation

- 6. Add 3.3ml of ammonium acetate protein precipitation solution, and vortex vigorously for 20 sec at high speed.
- 7. Centrifuge for 5 min at 3,000g. The precipitated proteins should form a tight, dark brown pellet.

DNA precipitation

- 8. Dispense 10ml isopropanol in to a clean 50ml centrifuge tube and add the supernatant from the previous step.
- 9.Mix by inverting gently 50 times until DNA is visible as thread or clump
- 10. Centrifuge for 5 min at 3,000g. Carefully discard the supernatant
- 11. Wash the DNA pellet by adding 10 ml of ethanol and centrifuge for 5 minutes at 3,000g and remove the supernatant.







DNA Hydration

- 12. Add 0.3-1.0 ml of DNA hydration buffer and vortex for 5 sec at medium speed to mix. Incubate at 65 degree C for 1 hour to dissolve DNA.
- 13. Incubate at RT overnight with gentle shaking.
- 14. The absorbance of the DNA at 260 nm and 280 nm should be measured using a quartz cuvette to assess purity as well as to known the approximate concentration of DNA